Characterization of the Structure and Dynamics of the HDV Ribozyme in Different Stages Along the Reaction Path

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ABSTRACT: The structure and dynamics of the hepatitis delta virus ribozyme (HDVr) are studied using molecular dynamics simulations in several stages along its catalytic reaction path, including reactant, activated precursor, and transition-state mimic and product states, departing from an initial structure based on the C75U mutant crystal structure (PDB: 1VC7). Results of five 350 ns molecular dynamics simulations reveal a spontaneous rotation of U-1 that leads to an in-line conformation and supports the role of protonated C75 as the general acid in the transition state. Our results provide rationale for the interpretation of several important experimental results and make experimentally testable predictions regarding the roles of key active site residues that are not obvious from any available crystal structures.

SECTION: Biophysical Chemistry

The hepatitis delta virus (HDV) ribozyme is a small catalytic RNA motif that is essential for viral replication during the HDV life cycle.1−4 Recently, HDV-like ribozymes have been found to be widely distributed in nature, including in human genes, where they likely play a variety of important biological roles.5 The HDV ribozyme (HDVr) catalysis reaction starts with an in-line nucleophilic attack of the U-1:O2 bond of the adjacent scissile phosphate, followed by cleavage of the P=O bond of G1 to produce a 2′,3′-cyclic phosphate and a 5′ hydroxy-terminus. Extensive structural and biochemical evidence suggests a catalytic mechanism involving acid−base catalysis; however, the detailed catalytic reaction mechanism of HDV ribozyme is still not resolved.

The first crystal structure of HDVr was reported in the product form with an overall fold consistent with mutagenesis and chemical probing studies of the solution conformation.7,8 In this product structure, C75:N3 is in a position to form a hydrogen bond with G1:O2′ (the leaving group); hence, it is reasonable to suggest that C75 acts as the general acid in phosphodiester bond cleavage reaction,9,10 a hypothesis that is well supported by inactivation of the ribozyme by C75 mutation and a variety of experimental approaches.9,11−13 Importantly, preevacuation of the 5′ leaving group by substitution with a 5′ bridging phosphorothioate renders the ribozyme insensitive to C75 mutation.10 Nevertheless, in the structures of the precleavage HDVr inactivated by C75U mutation and in the absence of Mg2+ ions, the active site has different arrangement in that U75 is poised to serve as the general base for cleavage reaction,14 an interpretation that has been supported in certain molecular dynamics studies.15−17

Although not absolutely required for catalysis,18 the presence of divalent metal ions at millimolar levels significantly enhances the HDVr reactivity.3,7,19−21 It is believed that there is a hydrated Mg2+ ion near the active site.9,14,22,23 This Mg2+ ion is likely to be involved in the HDVr catalytic reaction, as it has been shown that Co(NH3)6Cl3+ can compete with Mg2+ binding and inhibit HDVr activity.9,24 The active site Mg2+ ion has been shown to interact directly with critical active site residues,25 and modification of the linkage at the scissile phosphate can alter metal ion preference.26,27

Recently, Raman crystallographic experiments have determined that the pKa value of C75 is shifted toward neutrality in a Mg2+-dependent fashion12 and furthermore that protonation of C75 may be coupled to changes in inner-sphere coordination of a divalent metal ion binding24,25 Subsequent crystallographic26 and molecular dynamics29,30 studies have provided new information about the HDVr active site, and, in particular, the nature of metal ion binding at a site involving a G·U wobble at the scissile phosphate and a rare reverse G·U wobble base pair located near the active site. Nonetheless, the conformational events that lead to a catalytically active state where U-1 is poised for in-line attack are not well understood because this residue was not resolved in the recent crystallographic study but rather modeled based on the conformation of the inhibitor strand of the hammerhead ribozyme. Furthermore, there has been relatively little reported work on the
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Table 1. Summary of Simulations Reported in the Present Work with Their Abbreviations Used in the Text

<table>
<thead>
<tr>
<th>abbrev</th>
<th>state</th>
<th>C75</th>
</tr>
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<tbody>
<tr>
<td>RT-C75-Mg</td>
<td>reactant</td>
<td>0</td>
</tr>
<tr>
<td>dRT-C75-Mg</td>
<td>activated precursor</td>
<td>+</td>
</tr>
<tr>
<td>ETS</td>
<td>early TS mimic</td>
<td>+</td>
</tr>
<tr>
<td>LTS</td>
<td>late TS mimic</td>
<td>+</td>
</tr>
<tr>
<td>Prod</td>
<td>product</td>
<td>0</td>
</tr>
</tbody>
</table>

"TS" refers to "transition state" and "0" and "+" refer to neutral and protonated (at the N3 position) C75. Systems were solvated in a 60 × 60 × 120 Å box of TIP3P waters and an ion atmosphere corresponding to a 0.14 M bulk NaCl concentration, with residue C41 protonated. All simulations were carried out in the NPT ensemble at 1 atm and 298 K under periodic boundary conditions and using the smooth particle mesh Ewald method for calculation of electrostatic interactions. For each system, 10 ns of water/ion equilibration, followed by an additional 10 ns of solute equilibration were performed, followed by 350 ns of production simulation, the last 300 ns of which was used for analysis. Simulations were performed with the NAMD simulation package (version 2.7b3) using the AMBER parm99 force field with the α/γ corrections for nucleic acids.

HDVr structure and dynamics in different key stages along the catalytic reaction path.

The present study examines results from a series of molecular dynamics simulations of HDVr in different stages along the reaction path. Whereas previous simulation studies on HDVr focused on only the reactant states or the product state, here we report results from a series of five 350 ns molecular dynamics simulations of HDV ribozyme (summarized in Table 1), the first focusing on different points along the reaction coordinates: (1) the reactant state (RT) with neutral C75 (C75\(^+\)), (2) the activated precursor state with the nucleophile (U-1:O\(_2\)) deprotonated (dRT) and C75 protonated (C75\(^+\)), (3) the early transition-state mimic (ETS), (4) the late transition-state mimic (LTS), and (5) the product state (Prod). The ETS and LTS transition-state mimics are formed by defining new chemical bonds between the nucleophile O\(_2\) and the active center P atom, with different bond lengths derived from high-level ab initio quantum calculations. The same protocol has been previously utilized in various simulation work of hammerhead ribozymes, where significant differences were observed in the ETS and LTS simulation results. The unrefined starting structures were based on the 2.45 Å crystal structure of the C75U mutant (PDB: 1VC7) which contains positions for the U-1 residue but that differs significantly from crystallographic data for the product structure and from recent crystallographic data of HDVr bound to an inhibitor RNA containing a deoxynucleotide at the cleavage site. In this structure, a metal ion is at the position between U75 and G25 and is directly coordinated to U75:O\(_4\), which cannot occur in the wild type. Hence in our simulations the active Mg\(^{2+}\) ion was initially placed to be bound to G1:N7, in accord with the position suggested by Chen et al. In the first 10 ns of simulation, the Mg\(^{2+}\)–G1:N7 distance was restrained to 2.0 Å. Initial simulations with the active-site metal ion placed at the original position of the C75U crystal structure indicated that with the native C75 the metal ion does not bind stably at this position. Recent crystallographic structure of the inhibited reactant could be an alternative choice of the starting structure; however, the key nucleophile residue U-1 is missing in this structure and hence it was not selected here. The full details of the simulation protocol are provided in the Supporting Information.

The focus of this work is to present structure models in different stages of the HDVr reaction path. These simulation-derived models outline critical residues and their interactions, as shown in Figure 1. Different stages are shown: Figure 1A: the neutral reactant state (RT-C75-Mg); Figure 1B: the precursor state with the nucleophile deprotonated and C75 protonated (dRT-C75-Mg); Figure 1C: the early transition state mimic (ETS); Figure 1D: the late transition state mimic (LTS); and Figure 1E: the product state (Prod). Statistical analyses were performed for the key geometry indexes, including bond distances, angles, dihedrals, and hydrogen bonds. All indexes reported in the following sections were calculated from the last 300 ns out of the total 350 ns of trajectory for each simulation with a sampling rate of 100 ps. The hydrogen bonds are defined as formed when the distance between the donor and the acceptor is <3.5 Å and the angle is >150°. The H-bond is reported in terms of the percentage of the number of snapshots with formed hydrogen bonds compared with the total number of snapshots of each trajectory. The time series of mentioned H-bond distances are shown in Figure S1 of the Supporting Information.

In the neutral reactant state, G2:O\(_2\) positions U-1:O\(_2\) for general base activation by the active site Mg\(^{2+}\) ion. Substitution of sulfur at the G2:O\(_2\) position has a significant effect on HDVr activity that cannot be rescued by thiophilic metals, and as yet, the origin of this effect remains unclear. In the RT-C75-Mg simulation (Figure 1A), G2:O\(_2\) forms a hydrogen bond with the 1:O\(_2\) nucleophile (72% in a 300 ns trajectory). The interaction between G2:O\(_2\) and U-1:O\(_2\) is further facilitated by a Mg\(^{2+}\)–mediated water bridge involving two inner sphere water molecules (Figure 1A).

The C7s position is held near the active site by the H-bond between C75:N\(_4\) and the scissile phosphate G1:O\(_2\) (45%). Therefore, C75 is not in a position near the leaving group to

Figure 1. Graphic summary of the simulation results. Shown are representative snapshots from simulations listed in Table 1: (A) the neutral reactant state, (B) the precursor state with the nucleophile deprotonated and C75 protonated, (C) the early transition state mimic, (D) the late transition state mimic, (E) the product state, and (F) the crystal product structure (PDB ID: 1CX0).
act as a general acid, which is consistent with the precleavage crystal structure and other simulation results. However, the mechanisms cannot be unambiguously defined based on structural evidence corresponding to a single point along the reaction coordinate alone. In this case, it is particularly precarious to assume that C75 does not act as the general acid, given the growing body of experimental evidence. Furthermore, on the basis of its position in the precleavage state and the results from the RT-C75-Mg simulation, rearrangement of active site interactions is needed for C75 to participate in this catalytic mode.

In the activated precursor state, A77, the active site Mg$^{2+}$ and C75, collectively hold the in-line conformation formed after a rigid rotation of U-1. A representative snapshot from the simulation of the activated precursor state with the nucleophile deprotonated and C75 protonated (dRT-C75-Mg) is shown in Figure 1B. During the simulations, the U-1 residue spontaneously undergoes a rigid rotation and reaches a near-in-line conformation around 20 ns, with the O-P-S-O angle around 140°, rotates back to about 90° around 130 ns, and then rotates back to above 140° at around 200 ns. The in-line angle is kept around 140 degrees after 200 ns (average 138°, with maximum 162°). This observation shows that U-1 is able to adopt multiple conformations, consistent with recent crystallographic data of an inhibited precleavage structure where the electron density of U-1 was observed to be disordered. After 200 ns, the in-line conformation of U-1 is stabilized by a new H-bond between A77:N3 and the nucleophile (U-1-O2P) (38%), between C75:N4 and G1:O2P (30%), and between C75:N4 and G1:O2P (56%). The A77:N3-U1-O2P H-bond interaction is intriguing in that it provides a rationale for the hitherto unexplained importance of the exocyclic NH$_2$ group of A77 identified through mutagenesis experiments. The active site Mg$^{2+}$ is directly bound to G1:N7 (2.25 Å) and also bound to C75:O2 through a water molecule (5.20 Å). These interactions are consistent with experimental evidence, suggesting that a previously unobserved hydrated magnesium ion interacts with G1:N7 as well as the observation that the reactivity of the HDVr is reduced 28 fold when C75 is mutated to deoxy-C75.

In the early transition-state mimic simulation, U-1 forms a canonical WC pair with A78, and C75$^+$ forms strong H-bond with the leaving group. A dramatic change in the base pair hydrogen bonding occurs in the early transition-state mimic simulation, whereby a WC base pair forms between A78 and U-1 and is shown in Figure 1C. The occupancy of the H-bond between A78:N4 and U-1:O4 is 86%, whereas that between A78:N4 and U-1:N4 is 76%. This WC pair provides a rationale for the importance of the identity of A78. As well as the nucleobase preference of the −1 position and suggests that experiments involving correlated mutations in the 78 and −1 positions may provide further insight into the importance of this interaction, as it has been suggested that the U-1 preference can be altered under different conditions.

In the ETS, the protonated C75$^+$ moves to a position where it is available to act as the general acid and is held in place by a strong H-bond between C75:N4 (the exocyclic amine) and G1:O2P (84%). At the same time, the exocyclic amine of C75 forms an additional hydrogen bonding interaction with C22:O2P (57%), which helps to hold its position. The H-bond between C75:N4 and C22:O2P was observed crystallographically in the postcleavage structure and in a recent precleavage structure but not in the precleavage structure. The exocyclic amine of C75 (N4) thus appears to play an important role in maintaining the proper active site fold near the transition state, in agreement with conclusions from experiments that investigated the incorporation of 6-azauroidine into the genomic HDV active site. The Mg$^{2+}$ loses its direct coordination with G1:N7 and moves slightly away from the active site center when C75$^+$ moves toward G1:O2P, which is consistent with reported anticooperative binding of the Mg$^{2+}$ and C75 protonation.

In the late transition state, the position of the general acid is maintained by H-bond interactions between A77:N3 and G1:O2P and C75:N4 and C22:O2P. In the late transition-state mimic, cleavage of the P–OS$^-$ bond has progressed and there is an accumulation of negative charge at the G1:O2P position, leading to slight changes in the active site hydrogen bonding (Figure 1D). In the LTS, C75 has a similar position compared with the ETS, and the strong H-bond between C75:N4 and G1:O2P is maintained (65%). However, the WC pair between A78 and U-1, formed in the ETS is broken. Instead, because of the shift in the position of the reactive phosphate group, the exocyclic amine of A77: N4 now forms a strong H-bond with G1:O2P (40%). The H-bond between C75:N4 and C22:O2P is more pronounced in LTS (86%) than in ETS (57%). The Mg$^{2+}$ further loses both direct and indirect coordination interactions with G1:N7 and C75:O2P and exits from the active site as the reaction proceeds to a late stage. This ejection of a metal from the HDV active site in the late stages of the reaction has been previously inferred from crystallographic data.

In the product state, the Mg$^{2+}$ exits the active site and the simulation converges toward the crystal structure of the product complex. In our simulation of the product state, A77 and A78 lose all H-bond interactions with the substrate because U-1 no longer exists in the active site, as shown in Figure 1E. The H-bond between C75:N4 and G1:O2P, reported in the postcleavage structure, is formed (56%) as well as the H-bond between C75: N4 and C22:O2P (43%). The strong H-bond between C75:N4 and G1:O2P observed in other stages no longer exists because C75 is now in its neutral state, having donated its proton to the O2P leaving group. Our simulation of the product state converges reasonably closely to that of the crystallographic data of the product structure, with the exception of formation of a rare reverse G-U wobble base pair, also observed in a recent crystallographic structure of HDVr bound to an inhibitor RNA containing a deoxynucleotide at the cleavage site. This reverse wobble may be further stabilized by divalent metal ion binding.

A key point is that our simulations of the product state were initiated not from the product crystallographic structure but rather a precleavage structure of the C75U mutant. Therefore, the observation that despite beginning with a distinct geometry along the reaction coordinate our simulations converge in both structure and hydrogen bonding pattern very closely to that of the postcleavage structure, shown in Figures 1E,F and 2, provides an important internal check on reliability of our simulations.

The details of the catalytic mechanism of HDVr have been the topic of considerable discussion and debate, originating from varying mechanistic interpretations derived from crystallographic data and biochemical experiments. Of particular focus was the crystallographic structure of the C75U mutant in the precleavage state that suggested the role of C75 as a general base rather than general acid, as was inferred by previous crystallographic data of the native product. It was for this reason that we used this structure of the former as a departure point for our simulations of the native HDV at several stages along the reaction coordinate.
Our results suggest that the position of C75, which is initially close to the U-1:O2·- in the reactant state, prefers to adopt a hydrogen bonding interaction with the G1:O2p, whereas the nucleophile interacts with G2:O2p and a hydrated Mg2+ ion through a metal-mediated water bridge. These results do not support the role of C75 as that of a general base. Although the crystallographic structure of the C75U mutant was not in an in-line conformation required for nucleophilic attack, in the activated precursor simulation, U-1 reorients so as to form an in-line conformation that is stabilized by hydrogen bond interactions with A77.

The simulations of the transition state mimics indicate that protonated C75 adopts an orientation where it is poised to act as a general acid, acquiring interactions with the scissile phosphate and G1:O2p leaving group and being held in place, in part, by a hydrogen bond between the exocyclic amine of C75 and a nonbridge phosphoryl oxygen of C22 that supports a role for hydrogen bonding in the catalytic cycle. The active site is defined as the collection of G1, G2, G3, C75, A77, and A78. Data are shown every 100 ps, and the smooth solid lines along the data curves are the window-averaged results with window size = 10.

Furthermore, on the basis of this inhibited reactant structure and MD simulations, an alternate metal binding site near the G25/U20 pair has been proposed, where the G25·U20 reverse wobble base pair provides an environment for Mg2+ to bind to G25:N7 and nearby active site residues. In our simulations, G25 is in the anti conformation and its N7 is not facing the active site hence G25:N7 cannot provide such binding environment.

In all of our simulations reported here (total >1.5 μs), no G25:U20 reverse wobble base pair has been observed, consistent with a previously reported MD studies. Therefore, simulations with different G25 conformations may be needed to explore further the Mg2+ binding site near G25 and its relationship with the binding site we proposed in the present work. Experimentally, it would be of great interest to examine the HDVr activity with chemical modifications at the G25 position, including an N7 deaza modification to eliminate binding of Mg2+ to N7 or an 8-bromo substitution to favor the syn conformation required by the reverse wobble pair.

In conclusion, we present a set of extended molecular dynamics simulations for HDVr in different stages along the reaction path and characterize a conformational transition of U-1 into an in-line active conformation in the activated precursor state. Our simulations support the role of C75 as the general acid and identify several key residue interactions in different stages of the reaction. Our results provide explanations for the observed importance of several active site residues and suggest specific hypotheses that can be experimentally tested. Although simulations starting with other crystal structures may further explore different possible active site conformations, this work provides a departure point for further investigations into the catalytic chemical steps of the HDVr mechanism.

**ASSOCIATED CONTENT**

Supporting Information. Additional computational details. This material is available free of charge via the Internet at http://pubs.acs.org.

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